

Selection of coffee progenies with multiple resistance to biotic agents

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Introduction

Among the pathogens that affect coffee growing, the root knot nematodes (*Meloidogyne spp.*), the coffee rust (*Hemileia vastatrix*) and the bacterial halo blight (*Pseudomonas syringae pv. garcae*) pose a great threat to coffee production. The pyramiding of biotic stress resistance genes in highly productive cultivars has been one of the main objectives of *Coffea arabica* breeding programs. In this sense, the objective of this work was to select coffee trees with multiple resistance to root-knot nematodes, coffee rust and bacterial halo blight, through the identification of plants with multiple resistance to biotic agents in early generations of the breeding program conducted by the Agronomic Institute of Campinas.

Results/Discussion

Of 597 F₂ coffee plants inoculated with *P. syringae pv. garcae*, 372 proved to be resistant to the pathogen. As for the coffee rust, only 18 plants out of 355 tested were susceptible to *H. vastatrix* race II. Plants with resistance to coffee rust and bacterial halo blight were further evaluated for resistance to root knot nematodes. At the end of the experimental period it was possible to select 35 plants with high levels of root knot resistance and, consequently, with simultaneous resistance to all biotic agents studied (Table 1).

Table 1. Phenotyping of F₂ coffee populations in relation to the response to infection by the *P. syringae pv. garcae*, *Hemileia vastatrix* and *Meloidogyne spp.*

Population	<i>Pseudomonas syringae pv. garcae</i>			<i>Hemileia vastatrix</i>			<i>Meloidogyne spp.</i>		
	Scale of points			LD/TR			Damage index (DI)		
	R	S	n=	R	S	n=	R	S	n=
H 20393-1	78	44	122	79	75	122	73	0	73
H 20393-2	76	49	125	73	73	125	71	9	62
H 20393-3	74	44	118	70	69	118	71	3	68
H 20393-11	52	57	109	46	38	109	36	4	32
H 20393-13	92	31	123	87	82	123	82	19	63

Plant materials

- F₂ seeds originating from open pollination of five hybrids H 20393
- Parental of the hybrids H 20393:

Accession from Ethiopia (IAC 2036-6) × Cultivar (Sarchimor IAC 4933)
 (Font of resistance to bacterial halo blight and *Meloidogyne* spp.) (Font of resistance to coffee rust)

- Susceptibility standard: Cultivar IAC Mundo Novo 515-20

Multiple resistance evaluation

P. syringae pv. garcae

Experimental conditions
 (IBSBF 1197) 3x10⁸ CFU.ml⁻¹. Inoculation by sandpaper abrasion method (Rodrigues et al. 2017). The plants were kept in a damp chamber (humidity above 80%).

Evaluation methods

Evaluation 30 days after inoculation (Fig. 1A).
 - Scale from 0 to 5 points
 - Classification of resistant plants (0 and 1 points)
 - Chi-square test data analysis

H. vastatrix

Experimental conditions
 Uredospore suspension of *H. vastatrix* race II (0.025ml). Inoculation followed by incubation in the dark for 24h. (Esques & Toma-Braghini, 1981). (Fig. 1B).

Evaluation methods

- Lesion density (LD): scale from 0 to 9 points
 - Type of reaction (TR): scale from 0 to 4 points
 - Selection of resistant plants: LD and TR = 0 and 1

Meloidogyne spp.

Experimental conditions
 Plants inoculated with a mixture of *M. incognita*, *M. paranaensis* and *M. exigua* totaling 3000 eggs + J2/plant. Plants were kept in the green house conditions.

Evaluation methods

- Evaluation 180 days after inoculation (Fig. 1C).
 - Damage index (DI): scale from 0 to 5 points (Hussey & Jansen, 2002)
 - Classification of resistant plants: DI ≤ 2.

Materials/Methods

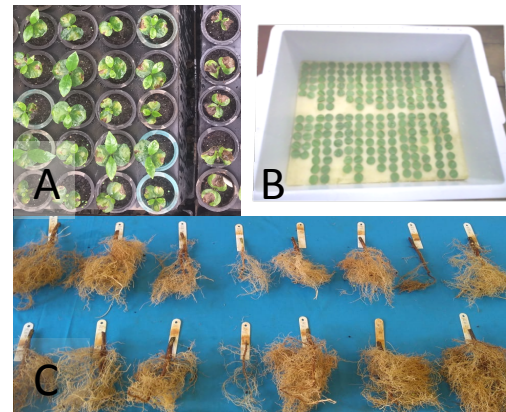


Figure 1. Part of the tests carried out for screening coffee trees with multiple resistance.
 A: Response of coffee trees to halo blight 30 days after inoculations with the IBSBF 1197 strain of *Pseudomonas syringae pv. garcae*.
 B: Detached disc test to evaluate resistance to race II of *Hemileia vastatrix*, the causal agent of coffee rust;
 C: Roots of coffee trees 180 days after inoculation of *Meloidogyne spp.* in mixture.

Conclusion/Perspectives

- 35 F₂ coffee trees were select with multiple resistance;
- Resistance to halo blight is probably qualitative in character, governed by few genes, as well as for individual races of *H. vastatrix*.

References

- Esques, A. B. & Toma-Braghini, M. FAO Plant Protection Bulletin, 1981, p. 56-66.
- Fatobene et al. Euphytica, 2017, p. 1-9.
- Rodrigues et al. Journal of Phytopathology, 2017, p. 105-114.